

Evaluation of Acute Toxicity Levels and Behavioral Changes to the Fry of Common Carp, *Cyprinus carpio* (Linnaeus, 1758) Under Exposure of a Pyrethroid Insecticide Cypermethrin

Chandan Sarkar, Nimai Chandra Saha*

Fishery and Ecotoxicology Research Laboratory, Department of Zoology, The University of Burdwan, Burdwan, Purba Barddhaman, West Bengal, India

Abstract

Toxicity of an insecticide Cypermethrin to the fry of common carp, Cyprinus carpio along with their behavioural changes was evaluated in the present study. The 24, 48, 72, and 96 h LC₅₀ values of Cypermethrin to the fry of Cyprinus carpio were 1.04, 0.92, 0.89, 0.79 ng/l, respectively. The mortality rate of C. carpio showed significant variation (p < .05) with the increasing concentrations at all exposure times. On the other hand, the mortality rate of C. carpio also varied significantly (p < .05) at all the doses with increasing exposure times (24, 48 and 72, and 96 h). The excess mucous secretion with increasing concentration of toxicant and exposure time was recorded in the exposed fish. The treated fish showed faster movement at lower doses but it was gradually reduced at higher doses at all exposure times. Wide opening of mouth and gills was observed at higher concentrations. Opercular movement of fish initially increases with the increasing concentration but decreases at higher concentrations. Similarly the opercular movement increases significantly (p < .05) at 96h of exposure at all the treatments.

Keywords: Behavioral change, cypermethrin, Cyprinus carpio, LC50, opercular movement

*Author for Correspondence E-mail: vcbunsaha@gmail.com

INTRODUCTION

The pollution of rivers, streams, and wetlands with chemical contaminants has become most serious problems of this century [1]. Among these harmful chemicals, insecticides occupy unique positions which are used regularly by man [2]. Insecticides applied directly to the field are washed away by flood and rainwater as runoff to adjacent wetlands and thereby alteration of physicochemical properties of water occurs [3]. Due to effectiveness and biodegradability, pyrethroids are very commonly used insecticides. Pyrethroids are synthetic analogues pyrethrins produced naturally by of Chrysanthemum cinerariaefolium, an ornamental plant [4]. Their 96 h LC_{50} for fish is below 10 µg/l. Thus, pyrethroids becomes highly toxic to fish and other aquatic organisms [4].

Cypermethrin (a-cyano-3-phenoxybenzyl ester of 2, 2-dimethyl-3-(2, 2-dichlorovinyl)-2-2dimethyl cyclopropane carboxylate) is extremely effective nonsystemic Type II synthetic pyrethroid. It is used in animal husbandry to resist ectoparasites and a wide range of other pests in agricultural field [5]. It acts by blocking sodium channels and affecting the function of GABA-receptors of nerve filaments [6]. The relatively slow metabolism and less efficiency to eliminate the compounds may cause the fish sensitivity to pyrethroids [7]. A large portion of world's food is come from fish source, so it is important to ensure the health of fishes [8].

The earlier studies indicate that the Cypermethrin is very toxic to various groups of fishes. According to Bradbury and Coats, 1989 [7], the 96-h LC₅₀ of Cypermethrin as $1.2 \mu g/L$ for brown trout (*Salmo trutta*), 0.5 $\mu g/L$ for rainbow trout (*Salmo gairdneri*), 0.4 $\mu g/L$ for *Scardinius erythropthalmus* and 2.2 $\mu g/L$ for

Tilapia nilotica. The lethal concentration (LC₅₀) value of cypermethrin was 0.063 mg/l for 96h of exposure in Clarias gariepinus (Burchell 1822) juvenile [2]. The acute toxicity value of Cypermethrin was found to be 4.0 µg/l for Labeo rohita fingerlings for 96 h [1]. According to Jones [9], in Brown trout (96 h) LC_{50} is 2.0-2.8 ppb, in Rainbow Trout (96 h) LC₅₀ is 0.82 ppb, in Bluegill Sunfish (96 h) LC_{50} is 1.78 ppm, in *Daphnia magna* LC_{50} is 0.26 ppb. In Poecilia reticulata (Peters) at 25° C, the 24, 48, 72, and 96h LC₅₀ values are 219.83, 202.72, 186.22, and 168.07 ppb, respectively [10]. According to Clark et al. (1989) [11], 96 h LC₅₀ of Cypermethrin for grass shrimp (Palaemonetes pugio) is 0.016 μ g/l. Collins et al (2006) [12] recorded the LC₅₀ of Cypermethrin to freshwater prawn Palaemonetes argentinus as 0.0031 and 0.0020 μ g/l for 24 and 96h, respectively. The reports on the toxicity of Cypermethrin to common carp (Cyprinus carpio) are scanty [7,13]. In Indian context, especially in West Bengal, the data of LC₅₀on fry of common carp during 24, 48, 72, and 96h, no earlier reports are found.

Therefore, the present study was aimed to determine the effect of acute toxicity of commonly used synthetic pyrethroid insecticide Cypermethrin on fry of common carp (*Cyprinus carpio*) and their behavioural changes.

MATERIALS AND METHODS

Fry of common carp (Cyprinus carpio) belonging to Order: Cypriniformes and family Cyprinidae (mean length 4.03 ± 0.70 cm, mean weight 2.35 ± 0.49 gm) was used as test animals in the bioassay. The specimens were obtained from local fish farm in early morning. They were transferred and stocked to laboratory immediately and allowed to acclimatize gradually to the test water for 72h inthere before experiment. The commercial grade of Cypermethrin (10% EC) was used as test chemical. To determine the acute toxicity of the test chemical, the static replacement bioassay method was conducted in 15 L glass aquarium each containing 10 L of unchlorinated tap water following the methods of earlier workers [14,15,16]. Fishes were randomly parcelled in glass aquaria (15 L) supplied with oxygenated

unchlorinated tap water water maintaining constant dissolved oxygen at 7.6 \pm 0.5 mg/L, temperature at $27 \pm 1^{\circ}$ C, pH at 7.13 ± 0.9 , total hardness 195 \pm 10 mg/L as CaCO₃ alkalinity 167 ± 11.89 mg/L and photoperiod at 14L/10D light: dark natural photoperiod. No food was supplied to the fishes for 24 h before and during the bioassay. Each experiment was conducted with four replicates with control. Each replicate was provided with ten fishes. Initially, rough range finding experiments were conducted to fix the dose range at which lethality of fish occurs. Finally, only the selected test concentrations of Cypermethrin were used to determine the LC₅₀ measure to *Cyprinus carpio* at 24, 48, 72, and 96 h of exposure. Fishes were considered lifeless when gill opercula and body movement ceased. The number of dead fish was noted at every 24 h of experiment. The dead fish were eliminated quickly from the aquaria to avoid any organic decomposition causing variation of dissolved oxygen. The 10% of the test water was changed by stock water at every 24 h of exposure time and desired quantity of test chemical was added quickly to test water to maintain a fixed concentration. To evaluate the effects of Cypermethrin on respiratory rate of fish, the mean opercular movements (number of movement/min/fish) were noted at every 24 h during the experiment. The behavioural changes like excess mucous secretion, movement, wide opening of mouth and gills were also recorded [17].

For statistical analysis, a statistical software, Probit program version 1.5 [18] were used to calculate the LC₅₀ values (95% confidence limit) using mean mortality of *Cyprinus carpio* after 24, 48, 72, and 96 h. The data of percentage of mortality and opercular movement were subjected to analysis of variance with the use of R-software [19] followed by Duncan's Multiple Range Test (DMRT) to find the significant variation among the mean values at different concentrations of Cypermethrin at different exposure times (24, 48, 72, and 96 h).

RESULTS AND DISCUSSION

The LC_{50} values (with 95% confidence interval) of Cypermethrin to *Cyprinus carpio* fry are shown in Table 1.

S STM JOURNALS

Table 1: LC₅₀ Values (With 95% ConfidenceInterval) of Cypermethrin to the CyprinusCarpio Fry at Different Times of Exposure(24, 48, 72, and 96 h).

Test	Concentration (ng/l)						
organism	24 h	48 h	72 h	96 h			
Cyprinus carpio	1.04	0.92	0.89	0.79			
	[0.97, 1.11]	[0.84, 0.99]	[0.79, 0.97]	[0.68, 0.86]			

The 24, 48, 72, and 96 h LC_{50} values of Cypermethrin to *Cyprinus carpio* are 1.04, 0.92, 0.89, 0.79 ng/l, respectively. The mortality rate of *C. carpio* showed significant

variation (p < .05) with the increasing concentrations at all exposure times (Table 2).

On the other hand, the mortality rate of *C*. *carpio* also varied significantly (p < .05) at all the doses with increasing exposure times (24, 48 and 72, and 96h). The excess mucous secretion with increasing concentration of toxicant and exposure time was recorded in the exposed fish. The treated fish showed faster movement at lower doses but it was gradually reduced at higher doses at all exposure times. Wide opening of mouth and gills was observed at higher concentrations (Table 3).

 Table 2: Mean Values (±SD) of % Mortality of Cyprinus Carpio Exposed to Different Concentrations of Cypermethrin at Different Times of Exposure (24, 48, 72, and 96 h).

Dose (ng/l)	Percentage of mortality of fish at different times of exposure (h)							
	24 h	48 h	72 h	96 h				
0.6	$00^{\mathrm{am}} \pm 0.00$	$00^{\mathrm{am}} \pm 0.00$	$10^{\mathrm{an}}\pm0.83$	$20^{ao}\pm0.50$				
0.7	$00^{am} \pm 0.43$	$20^{bn}\pm0.43$	$30^{bo}\pm0.50$	$40^{bp}\pm0.71$				
0.8	$20^{bm}\pm0.71$	$30^{cn} \pm 0.87$	$40^{\rm co}\pm 1.12$	$60^{cp} \pm 1.09$				
0.9	$20^{bm} \pm 0.43$	$50^{dn} \pm 0.50$	$50^{dn}\pm0.50$	$60^{co} \pm 0.43$				
1.0	$40^{\rm cm} \pm 0.50$	$60^{en} \pm 0.43$	$60^{en} \pm 0.43$	$70^{do}\pm0.43$				
1.1	$60^{dm} \pm 0.87$	$70^{\rm fn}\pm 0.71$	$70^{fn}\pm0.43$	$80^{\rm eo}\pm 0.71$				
1.2	$60^{dm} \pm 0.83$	$80^{\text{gn}} \pm 0.83$	$70^{\rm fo}\pm0.71$	$90^{\text{fp}}\pm0.83$				
1.3	$80^{\text{em}} \pm 0.83$	$90^{hn}\pm0.87$	$90^{\mathrm{gn}}\pm0.83$	$100^{\text{go}} \pm 0.43$				
1.4	$90^{\text{fm}} \pm 0.83$	$100^{\text{in}} \pm 0.00$	$100^{hn} \pm 0.00$	$100^{gn} \pm 0.00$				
1.5	$100^{\rm gm} \pm 0.50$	$100^{\text{im}} \pm 0.00$	$100^{\rm hm} \pm 0.00$	$100^{\text{gm}} \pm 0.00$				

Note. Mean values within columns indicated by different superscript letters (a-i) and mean values within rows indicated by different superscript letters (m-p) are significantly different (DMRT at 5% level).

Tuble 5. Demavioural Responses of Cyprinus Curpto.												
	Behavioural responses of fish at different times of exposure											
Dose (ng/l)	24 h		48 h		72 h		96 h					
	М	MS	WOMG	М	MS	WOMG	М	MS	WOMG	М	MS	WOMG
0.0	+++	-	-	+++	-	-	+++	-	-	+++	-	-
0.6	+++	-	_	+++	-	_	+++	-	-	++	1	+
0.7	+++	-	-	++	-	-	++	+	_	++	+	+
0.8	++	-	-	++	+	-	++	+	+	+	+	+
0.9	++	+	-	++	+	++	+	+	++	+	++	++
1.0	++	+	+	+	+	++	+	+	++	+	++	++
1.1	++	+	++	+	+	++	+	++	++	+	++	++
1.2	+	+	++	+	+	++	+	++	++	+	++	++
1.3	+	+	++	+	++	++	+	++	+++	-	++	++
1.4	+	+	++	+	++	+++	-	++	+++	Х	Х	Х
1.5	+	+	+++	X	Х	Х	Х	Х	Х	Х	Х	Х

Table 3: Behavioural Responses of Cyprinus Carpio.

Note. M = movement; MS = mucous secretion; WOMG = wide opening of mouth and gills; -: absent, +: mild, ++: moderate, +++: high, X = not recorded due to death (exposed to different concentrations of Cypermethrin at different times of exposure).

Daga ng/l	Opercular movement/minute of fish at different times of exposure								
Dose lig/l	24 h	48 h	72 h	96 h					
0.0	$85^{\rm eo}\pm 0.50$	$85^{\rm fo}\pm0.43$	$82^{en} \pm 0.43$	$72^{gm}\pm0.00$					
0.6	$90^{\rm fn}\pm 0.00$	$120^{ho}\pm0.50$	$90^{\rm fn}\pm 0.43$	$78^{hm}\pm0.43$					
0.7	$95^{\text{gm}}\pm0.43$	$120^{hn}\pm0.50$	$130^{ho}\pm0.50$	$95^{im}\pm0.50$					
0.8	$100^{hn}\pm0.50$	$110^{\rm go}\pm0.43$	$115^{\text{gp}}\pm0.50$	$60^{\rm fm}\pm 0.43$					
0.9	$84^{\rm eo}\pm 0.50$	$82^{en}\pm0.50$	$90^{\rm fp}\pm 0.50$	$55^{em}\pm0.50$					
1.0	$80^{do}\pm0.43$	$77^{dn}\pm0.43$	$80^{\rm fo}\pm0.43$	$54^{dm}\pm0.43$					
1.1	$65^{co} \pm 0.50$	$64^{cn} \pm 0.50$	$70^{cp} \pm 0.00$	$50^{cm}\pm0.43$					
1.2	$63^{bo}\pm0.50$	$60^{bn}\pm0.43$	$65^{bp} \pm 0.50$	$47^{bm}\pm0.50$					
1.3	$60^{ap} \pm 0.43$	$57^{ao} \pm 0.43$	$55^{an} \pm 0.50$	$45^{am} \pm 0.43$					
Note. Mean values within columns indicated by different superscript letters $(a-i)$ and mean values within rows indicated by different superscript letters $(m-p)$ are									

Table 4: Mean Values (±SD) of Opercular Movement/Minute of Cyprinus Carpio Exposed to Different Concentrations of Cypermethrin at Different Times of Exposure (24, 48, 72, and 96 h).

Opercular movement of fish initially increases with the increasing concentration but decreases at higher concentrations (Table 4).

significantly different (DMRT at 5% level).

Similarly, the opercular movement increases significantly (p < .05) with increasing exposure time upto 72 h but it decreases significantly (p < .05) at 96 h of exposure at all the treatments.

In the current study, the 24 and 96 h LC_{50} value (1.04 and 0.79 ng/l) corresponds to the LC_{50} of freshwater prawn Cypermethrin to Palaemonetes argentinus as 0.0031 and 0.0020µg/l for 24 and 96 h, respectively [12]. The 96h LC_{50} value of the present study (0.79 ng/l) is much lower than the 96h LC₅₀ value 1.2 µg/L for brown trout (Salmo trutta), 0.5 µg/L for rainbow trout (Salmo gairdneri), 0.4 µg/L for Scardinius erythropthalmus and, 2.2 µg/L for Tilapia nilotica [7], 0.063 mg/l for Clarias gariepinus (Burchell 1822) juvenile [2], 1.78 ppm in Bluegill Sunfish [9]. The 24, 48, 72, and 96h LC_{50} value of the present study (1.04, 0.92, 0.89, 0.79 ng/l, respectively) are also much lower than 24, 48, 72, and 96h LC₅₀ values of Poecilia reticulata (Peters) at 25°C (219.83, 202.72, 186.22, and 168.07 ppb, respectively) [10]. It indicates that the fry stage of Cyprinus carpio was comparatively more sensitive to Cypermethrin than brown trout (Salmo trutta), rainbow trout (Salmo gairdneri), Clarias gariepinus (Burchell 1822) juvenile, bluegill sunfish, and Poecilia reticulata (Peters).

The abnormal behaviors like excess mucous secretion, erratic movement, wide opening of mouth and gills observed in the current study were probably due to the enzymatic as well as ionic alteration in blood and tissues [20]. The excess mucous secretion of fish was probably for the dysfunction of endocrine gland especially pituitary gland over the integument for xenobiotic stress [21]. The abnormal behaviors of the treated fish may be due to adjust and adapt a compensatory mechanism to obtain energy to escape from stress due to toxicity of insecticide [22].

The results indicate that such low amount of Cypermethrin (0.7 ng/l) in aquatic ecosystem may have a significant effect on fry of *Cyprinus carpio* populations. Moreover, according to Kumaragura and Beamish [23], acute toxicity of synthetic pyrethroids to fish population was negatively correlated to temperature. Thus, the presence of pyrethroids in the aquatic ecosystem when water temperature decreased may increase the toxical influence of Cypermethrin on fish [24].

CONCLUSION

The results of the present experiment may provide supplement to the current knowledge on toxicity of Cypermethrin. In conclusion, Cypermethrin contamination is seriously harmful to aquatic ecosystems and this information should be taken into thought when this insecticide is applied in agriculture.



ACKNOWLEDGEMENT

The authors are grateful to the Head, Department of Zoology, The University of Burdwan for extending infrastructural facilities to carry out the work.

REFERENCES

- 1. Marigoudar SR, Ahmed RN, and David M. Cypermethrin induced respiratory and behavioural responses of the freshwater teleost, *Labeo rohita* (Hamilton), *Veterinarski Arhiv.* 2009; 79 (6), 583– 590p.
- 2. Ayoola SO, Ajani EK. Histopathological effects of Cypermethrin on juvenile African catfish (*Clarias gariepinus*). World J Biol Res. 2008; 1: 2p.
- Olufayo MO, Alade OH. (2012), Acute toxicity and histological changes in gills, liver and kidney of catfish, *Heterobranchus bidorsalis* exposed to Cypermethrin concentration, *African J Agri Res.* 2012; 7 (31): 4453–4459p
- 4. Dobsikova R, Velisek J, Wlasow T, Gomulka P, Svobodova Z, Novotny L. of Cypermethrin Effects on some haematological, biochemical and histopathological parameters of common (Cyprinus carpio carp L.). Neuroendocrinology Letters. 2006; 27: 91-95p.
- 5. Treasurer JW, Wadworth SL. (2004) Interspecific comparison of experimental and natural routes of *Lepeophtheirus salmonis* and *Caligus elongates* challenge and consequences for distribution of chalimus on salmonids and therapeutant screening. *Aquac Res.* 2004; 35 (8): 778– 783p.
- Roberts T, Hudson D. (1999); Metabolic pathway of agrochemicals: Part 2: Insecticides and fungicides. Cambridge, England: The Royal Society of Chemistry; 1999.
- Bradbury SP, Coats JR. Comparative toxicology of the pyrethroid insecticide. *Rev Environ Contam T.* 1989; 108: 133– 177p.
- Tripathi, G, Harsh S. (2002). Fenvalerateinduced macromolecular changes in the catfish *Clarias batrachus*. *J. Environ. Biol.* 2002; 23: 143–146p.

- 9. Jones DA. *Environmental* fate of *Cypermethrin* [Online]. Environmental Monitoring and Pest Management Department of Pesticide Regulation Sacramento. Available from https://www.cdpr.ca.gov/docs/emon/pubs/ fatememo/cyperm.pdf [Accessed on September 2018].
- Gautam PP, Gupta AK. Toxicity of cypermethrin to the juveniles of freshwater fish *Poecilia reticulata* (Peters) in relation to selected environmental variables. *Nat Prod Radiance*. 2008; 7 (4): 314–319p.
- 11. Clark JR, Goodman LR, Bortwick PW, Patrick JM, Cripe GM, Moody PH, More JC, Lores EM. Toxicity of pyrethroid to marine invertebrates and fish: A literature review and results with sediment-sorbed chemicals. *Environ Toxicol Chem*. 1989; 8: 393–401p.
- 12. Collins P Capello S. Cypermethrin toxicity to aquatic life: Bioassay for the freshwater prawn, *Palaemonetes argentines*. Arch Environ Contam Toxicol. 2006; 18: 10–14p.
- 13. Wang Y, Xiong L, Liu XP, Xie T, Wang K, Huang XQ, *et al.* Subacute toxicity of cypermethrin to carp. *J Agro-Environ Sci*; 2006, 25: 200–203p.
- Saha, NC, Bhunia F, Kaviraj A. Toxicity of phenol to fish and aquatic ecosystems. *Bulletin of Environ Contam Toxicol.* 63: 195–202p.
- American Public Health Association (APHA). Standard methods for the examination of water and waste water. Washington DC: American Public Health Association, American Water Works Association, Water Environment Federation; 2012.
- Sarkar C, Bej S, Saha NC. Acute toxicity of triazophos to common carp (*cyprinus carpio*) fry and their behavioural changes; *Paripex-Indian J Res.* 2016; 5 (6): 19–21p.
- Mukherjee D, Saha NC. Evaluation of acute toxicity levels and ethological responses under Tetrachlorocatechol exposure in common carp, Cyprinus carpio (Linnaeus). *Proceed Zoolog Soc*, Springer, 2013; 67 (2): 108–113p.
- US EPA. Probit program version 1.5. Cincinnati, Ohio: Ecological Monitoring Research Division, Environmental Monitoring Systems Laboratory, US Environmental Protection Agency; 1999.

- R Development Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2011.
- 20. Larsson, A., Bengtsson BE, Haux C. (1981). Disturbed ion balance in flounder, Platichthyesflesus L. exposed to sublethal levels of cadmium. *Aqua Toxicol*. 1981; 1 (1): 19–35p.
- 21. Pandey A, Kumar GK, Munshi JSD. Integumentary chromatophores and mucus glands of fish as indicator of heavy metal pollution. *J Freshwater Biol*. 1990; 2: 117– 121p.
- 22. Joshi PS. Impact of zinc sulphate on behavioural responses in the freshwater fish *Clarias batrachus* (Linn.). *Online Intl Interdisciplinary Res J.* 2011; 1 (2): 76–82p.
- 23. Kumaragura AK, Beamish FWH. Lethal toxicity of permethrin to rainbow trout,

Salmo gairdneri, in relation to body weight and temperature. *Water Res.* 1981; 15: 503–505p.

24. Moore A. Waring CP. The effect of a synthetic pyrethroid pesticide on some aspects of reproduction in Atlantic salmon (*Salmo salar* L.). *Aqua Toxicol*. 2001; 52: 1–12p.

Cite this Article

Chandan Sarkar, Nimai Chandra Saha. Evaluation of Acute Toxicity Levels and Behavioral Changes to the Fry of Common Carp, Cyprinus carpio (Linnaeus, 1758) Under Exposure of a Pyrethroid Insecticide Cypermethrin. *Research & Reviews: A Journal of Life Sciences*. 2018; 8(3): 102–107p.