

Hematobiochemical Alteration in Naturally Infected Cattle with Fasciola under Tropical Region

Anil Gattani¹*, Ajeet Kumar¹, G. D. Singh², Ramesh Tiwary², Anil Kumar², Arvind Kumar Das³, S. Samantaray¹

¹Department of Veterinary Biochemistry, Bihar Veterinary College, Patna, Bihar, India ²Department of Veterinary Clinics, Bihar Veterinary College, Patna, Bihar, India ³Department of Veterinary Medicine, Bihar Veterinary College, Patna, Bihar, India

Abstract

Fascioliasis is among one of the emerging and re-emerging disease which has zoonotic importance and spread by feco-oral route. The migrating fluke causes extensive liver damage results in economic burden to the animal farmer. In present investigation 20 cattle were used to evaluate the hemato-biochemical alteration in naturally acquired fasciola infection and without other diseases. Decreased Hemoglobin, Packed cell volume (PCV), Red blood cells (RBC) count whereas; an elevated level of Mean corpuscular volume (MCV) and Mean corpuscular hemoglobin (MCH) was observed in infected cattle. Hypoproteinemia, hypoalbunemia and hypoglycemia was recorded in infected cattle. The activities of serum enzyme Aspartate aminotransferase (AST), Alanine amino transferase (ALT), Gammaglutamyl tranferase (GGT) and Alkaline phosphatase (ALP) were increased in fasciola infection.

Keywords: Fasciola, hypoproteinemia, hypoalbuminemia, GGT

*Author for Correspondence E-mail: gattanianil@gmail.com

INTRODUCTION

The fascioliasis, a food borne zoonotic disease, that affects herbivores, preferably to cattle and sheep, and less frequently the man, which is caused by the parasite Fasciola along with Fasciola gigantica hepatica (F. gigantica) [1]. Because of the changes in the climate fascioliasis is emerging or rein many emerging countries, and its prevalence, intensity geographic and distribution are increasing [2]. In domestic fascioliasis include annual losses of more than US\$3000 million to livestock production worldwide through livestock mortality and by decreased meat and milk productions, decreased female fertility and increased veterinary costs [3]. These flukes reside and graze on the mucosa of the bile ducts and liver parenchyma resulting in massive liver damage [4]. Liver performs a number of essential function and play a very vital role in vertebrates and hepatic damage has a variety of detrimental effects and the metabolic functions of the liver are gradually reduced in fascioliasis. Various biochemical attributes viz. blood glucose, creatinine, urea and serum proteins while the activity of certain serum enzymes such as; AST, ALT and ALP, GGT which increase following hepatic injury, can be measured for hepatic functions [5]. Very few studies on hemato-biochemical alterations due to naturally acquired fascioliasis in cattle have been documented. Therefore, the biochemical alterations in cattle due to bovine fascioliasis using cattle with naturally acquired fasciola infection and without other diseases using hemato-biochemical diagnostic procedure that are reliable has been carried out.

MATERIAL AND METHOD

The present experiment was conducted at Bihar Veterinary College, Patna on 20 noninfected cattle and 20 cattle with naturally acquired bovine fascioliasis and no other disease. Blood samples for both hematological (with anticoagulant) and serum biochemical (without anticoagulant) studies were collected from cattle from Sept 2013 to Oct 2014. Serum samples were harvested using the standard procedure for biochemical studies. The enzymatic estimations were done within 24 h of collection. The hematology was carried out using standard procedures. The erythrocytes indices like mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) was calculated using the standard formulae. The serum biochemical parameters including serum glucose, protein, albumin, blood urea, chloride, calcium, phosphorus, Alanine aminotrasferase (ALT), Aspartatate aminotransferase (AST) were estimated using kits supplied by Span Diagnostics on Spectrophotometer. The data were subjected to statistical analysis by using SPPSS version 17 [6].

RESULTS AND DISCUSSION

In the present investigation, the following findings for the clinical condition of the cattle were assessed: pale mucus membrane, anemia, weakness, frequent diarrhea, emaciation, anorexia, intermittent fever, weight loss, swelling at brisket region. Statistically significant low (P<0.05) concentration were seen in the hemoglobin, PCV, Total erythrocyte count (TEC) whereas; a significant increase was seen in MCV and MCH along with a non-significant effect in MCHC (P>0.05). In the analysis of serum of the infected animals, when compared with controls, it was found that there were statistically significant decrease in glucose, total protein, albumin and significant increase in the mean serum activity of aspartate aminotransferase, alanine aminotransferase, ytransferase, alkaline glutamyl and phosphatase. Additionally, changes in hematobiochemical parameters in the infected animals and control group are given in Table 1.

Table 1: Hematobiochemical Values	in
Control and Fasciola Infested Cattle	2.

S. No.	Parameter	Control (n= 20)	Infested (n= 20)
1.	TEC	6.61 ± 0.22	4.33 ± 0.15
2.	Hb	10.22 ± 0.3	5.99 ± 0.18
3.	PCV	31.46 ± 1.01	18.67 ± 0.58
4.	MCV	40.28 ± 1.31	50.01 ± 0.79
5.	MCH	13.71 ± 0.61	15.67 ± 0.33
6.	MCHC	32.44 ± 0.25	32.34 ± 0.20
7.	Glucose	62.29 ± 2.21	45.82 ± 1.40
8.	Total Protein	6.86 ± 0.08	5.37 ± 0.17
9.	Albumin	3.52 ± 0.04	2.47 ± 0.07
10.	AST	102.91 ± 3.70	220.01 ± 21.21
11.	ALT	32.89 ± 1.95	100.44 ± 7.98
12.	GGT	14.09 ± 0.78	43.97 ± 2.65
13.	ALP	83.26 ± 7.79	231.83 ± 12.17

In this study, the biochemical parameters of non-infected cattle were found to be within the reference ranges for normal cattle [7, 8]. Statistical analysis revealed a significantly (p<0.05) lower concentration of Hb, PCV, RBC in infected cattle than the control group. On the other end a significant (p<0.05) higher values of MCV and MCH were observed in infected group. However, a non-significant difference was observed for MCHC. Present findings are similar to other researchers in cattle [9,10] and in sheep [11]. The reduction in RBC counts, Hb and PCV in present investigation may be due to the acute loss of blood caused by the flukes or extensive loss of blood into bile duct due to the large amounts of flukes present in the liver. Severe anemia may occur because of chronic liver inflammation, which causes suppression in erythropoesis [7, 12, 13]. Hemoglobin and PCV may be a useful tool in predicting the extent of fluke load and in indicating the prognosis of infected sheep [14]. On the other hand, significant increase was obtained in MCV and MCH of the infected cattle, when compared with those of the control group. Similar higher MCV in the infected animals were reported earlier [10, 15].

Glucose concentration in present study was significantly lower (p < 0.05) in the infected cattle the control than group. The hypoglycaemia may be due to the disturbance in gluconeo-genesis, which resulted from hepatic disorder [16], elevation of the ketone bodies from gastroenteritis could result in depression in blood glucose [17], besides the depression of the hepatic gluconeogenic pathways and decrease in voluntary feed intake by the infected cattle [18]. In present study hypoproteinemia and hypoalbuminemia was observed in fasciola infected cattle. Total serum protein and albumin were significantly lower (p<0.05) in infected groups. This in agreement with the report of Anderson [19] in cattle infected with F. hepatica. Because of extensive liver damage the synthetic function of liver is hampered and may lead to reduced synthesis of albumin. The infestation of fasciola may produce cholangitis, biliary obstruction, destruction and fibrosis of hepatic tissue and anaemia [8]. Hypoproteinaemia may be due to infection of the liver, which produce destruction of liver parenchyma



resulting in drastic alteration in protein values [20]. Activities of serum enzymes AST, ALT, GGT and ALP were significantly higher (p< 0.05) in the infected group. The results of present investigation are in concurrence with published work [21, 22]. An elevation in serum enzymatic activities may also be attributed to the degenerative changes and cirrhosis of the liver cells and enlargement of gall bladder. Moreover, the cellular changes from parasitism increase the permeability of the hepatic cells and in turn result in the release of the enzymes into the serum. Since ALP is known to be excreted via the bile duct, its elevation may have synchronized with the arrival of the flukes to the bile duct. Elevation in GGT levels was an indicator of chronic changes, cholestasis and epithelial damage in bile ducts caused by presence of adult flukes in biliary tract [20, 23, 24]. However, the GGT activity increased in infected llamas due to hepatic toxicity and necrosis [25]. So it may be ascribed that the elevations in serum AST, ALT, GGT and ALP activities were sensitive indicators of hepatic cell damage and hepatic dysfunction in fascioliasis and that the hepatic damage was hepatobiliary.

CONCLUSION

This present work led to conclude that measuring the hemato-biochemical parameters could be useful in early diagnosis and prognosis of bovine fascioliasis.

REFERENCES

- 1. Morphew MR, Wright HA, LaCourse EJ, et al. Towards Delineating Functions within the Fasciola Secreted Cathepsin 1 Protease Family by Integrating *in vivo* based Sub-proteomics and Phylogenetics. *Plos Nglec Trop Dis.* 2011; 5: e937p.
- Mas-Coma S. Human Fascioliasis. In: World Health Organization – Waterborne Zoonoses, Identification, Causes and Control. London, IWA Publishing, Alliance House. 2004.
- CalleÂjaa C, Bigota CK, Eeckhouttea P, et al., Comparison of Hepatic and Renal Drug-metabolising Enzyme Activities in Sheep given Single or Two-fold Challenge Infections with Fasciola Hepatica. Int J Parasitol. 2000; 30: 953–58p.

- 4. Shaikh AA, Bilqees FM, Khan MM. Histopathology of the Liver of Cow Due to Fasciola Gigantic Infection. *Proc. Parasitol.* 2005; 40: 17–24p.
- Craig AM, Pearson, EG, Rowe K. Serum Bile-acid Concentrations in Clinically Normal Cattle - Comparison by Type, Age, and Stage of Lactation. Am J Vet Res. 1992; 53(10): 1784–86p.
- 6. Snedecor GW, Cochran WG. *Statistical Methods.* 7th Ed. Oxford and IBH Publishing Co., Calcutta. 1980.
- Kaneko JJ, Harvey JW, Bruss ML. *Clinical Biochemistry of Domestic Animals.* 5th Ed., Academic Press. London. 1997.
- 8. Blood DC, Radostits OM, Arundel JH, *et al. Veterinary Medicine*. 7th Ed. Bailliere Tindall, London, U.K. 1989.
- 9. Molina EC, Lozano SP, Barraca AP. The Relationship between Haematological Indices, Serum Gamma-glutamyl Transferase and Glutamate Dehydrogenase, Visual Hepatic Damage and Worm Burden in Cattle Infected with *Fasciola gigantica. J Helminthol.* 2006; 80(3): 277–79p.
- Haroun EM, Hussein MF. Clinicopathological Studies on Naturallyoccurring Bovine Fascioliasis in the Sudan. J Helminthol. 1975; 49(3): 143– 52p.
- Doaa FT, Soliman EK, Abd EL- Khalek TMM. Effect of Fascioliasis on Hematological, Serum Biochemical and Histopathological Changes in Sheep. *Egypt J. Sheep Goat Sci.* 2007; 2(2): 15– 34p.
- Kramer JW. Normal Hematology of Cattle, Sheep and Goats. In: Feldman BF, Zinkl JG and Jain NC (Eds) Schalm's Veterinary Haematology, 5th Ed. Lippincott Williams and Wilkins, Philadelphia. 2000.
- 13. Lotfy HS, Mahmoud SM, Abdel-Gawad MA. Some Studies on Fascioliasis in Mid-Egypt. *Agr Res.* 2003; 81(2): 209–27p.
- 14. Hawkins CD. The Use of Haemoglobin, Packed-cell Volume and Serum Sorbitol Dehydrogenase as Indicators of the Development of Fascioliasis in Sheep. *Vet Parasitol.* 1984; 15: 125–33p.

- Sykes AR, Coop RL, Rushton B. Chronic Subclinical Fascioliasis in Sheep: Effects on Food Intake, Food Utilisation and Blood Constituents. *Res Vet Sci.* 1980; 28: 63–70p.
- Soulsby EJL. Helminthes, Arthropods and Protozoa of Domesticated Animals, 7th Ed. Balliere Tindall, London, UK. 1982.
- 17. Duncan JR, Prasse KW, Mahaffey EA. *Veterinary Laboratory Medicine*. 3rd Ed. Iowa State University Press, USA. 1994.
- Phiri IK, Phiri AM, Harrison LJS. The Serum Glucose and β-hydroxybutyrate Levels in Sheep with Experimental Fasciola Hepatica and Fasciola Gigantica Infection. *Vet Parasitol.* 2007; 143: 287– 93p.
- 19. Anderson PH, Berrett S, Brush PJ, *et al.* Biochemical Indicators of Liver Injury in Calves with Experimental Fascioliasis. *Vet Rec.* 1977; 15: 43–45p.
- Matanović K, Severin K, Martinković F, et al. Hematological and Biochemical Changes in Organically Farmed Sheep Naturally Infected with Fasciola Hepatica. J Parasitol Res. 2007; 101(6): 1463–65p.
- El-Aziz MZA, Emara SA, Salem FS. Clinicopathological Studies on Fascioliasis among Sheep in Giza Province. *Egypt J Vet Sci.* 2002; 36: 75– 86p.

- 22. Ahmed MI, Ambali AG, Baba SS. Hematological and Biochemical Responses of Balami Sheep to Experimental Fasciola Gigantica Infection. *J Food, Agr Environ* 2006; 4(2): 71–74p.
- 23. Ferre I, Lopez P, Gonzalo-Orden M, *et al.* The Effects of Subclinical Fascioliasis on Hepatic Secretory Function in Sheep. *Parasitol Res.* 1995; 8: 127p.
- 24. Taleb DF, Soliman EK, el- Khalek A. Effect of Fascioliasis on Hematological, Serum Biochemical and Histopathological Changes in Sheep. *Egypt J Sheep Goat Sci.* 2007; 2: 15–34p.
- 25. Duff JP, Maxwell AJ, Calxton JR. Chronic and Fatal fascioliasis in Llamas in the UK. *Vet Rec.* 1999; 145: 315–16p.

Cite this Article

Anil Gattani, Ajeet Kumar, Singh G. D., *et al.*; Hematobiochemical Alteration in Naturally Infected Cattle with Fasciola under Tropical Region; Research & Reviews: *Journal of Veterinary Science and Technology*. 2015; 4(1): 20–23p.